

Copy Number Variants: detection and analysis

Manuel Ferreira & Shaun Purcell

Boulder, 2009

Large chromosomal rearrangements can cause sporadic disease

Down syndrome

Duchenne Muscular Dystrophy (DMD)

DiGeorge-Velo cardiofacial syndrome (VCFS)

...

Large-Scale Copy Number Polymorphism in the Human Genome

Jonathan Sebat,¹ B. Lakshmi,¹ Jennifer Troge,¹ Joan Alexander,¹ Janet Young,² Pär Lundin,³ Susanne Månér,³ Hillary Massa,² Megan Walker,² Maoyen Chi,¹ Nicholas Navin,¹ Robert Lucito,¹ John Healy,¹ James Hicks,¹ Kenny Ye,⁴ Andrew Reiner,¹ T. Conrad Gilliam,⁵ Barbara Trask,² Nick Patterson,⁶ Anders Zetterberg,³ Michael Wigler^{1*}

Detection of large-scale variation in the human genome

A John Iafrate^{1,2}, Lars Feuk³, Miguel N Rivera^{1,2}, Marc L Listewnik¹, Patricia K Donahoe^{2,4}, Ying Qi³, Stephen W Scherer^{3,5} & Charles Lee^{1,2,5}

We identified 255 loci across the human genome that contain genomic imbalances among unrelated individuals. Twenty-four variants are present in >10% of the individuals that we examined. Half of these regions overlap with genes, and many coincide with segmental duplications or gaps in the human genome assembly. This previously unappreciated heterogeneity may underlie certain human phenotypic variation and susceptibility to disease and argues for a more dynamic human genome structure.

Iafrate et al 2004 Nat Genet 36: 949

which large duplications and deletions contribute to human genetic diversity is unknown. Here, we show that large-scale copy number (CNPs) (about 100 kilobases and greater) contribute substantially to genetic variation between normal humans. Representational oligonucleotide microarray analysis of 20 individuals revealed a total of 221 copy number differences, including 76 unique CNPs. On average, individuals differed by 11 CNPs, and the average length of a CNP interval was 465 kilobases. We observed copy number variation of 70 different genes within CNP intervals, including genes with diverse biological function, regulation of cell growth, regulation of metabolism, and several genes known to be associated with disease.

Sebat et al 2004 Science 305: 525

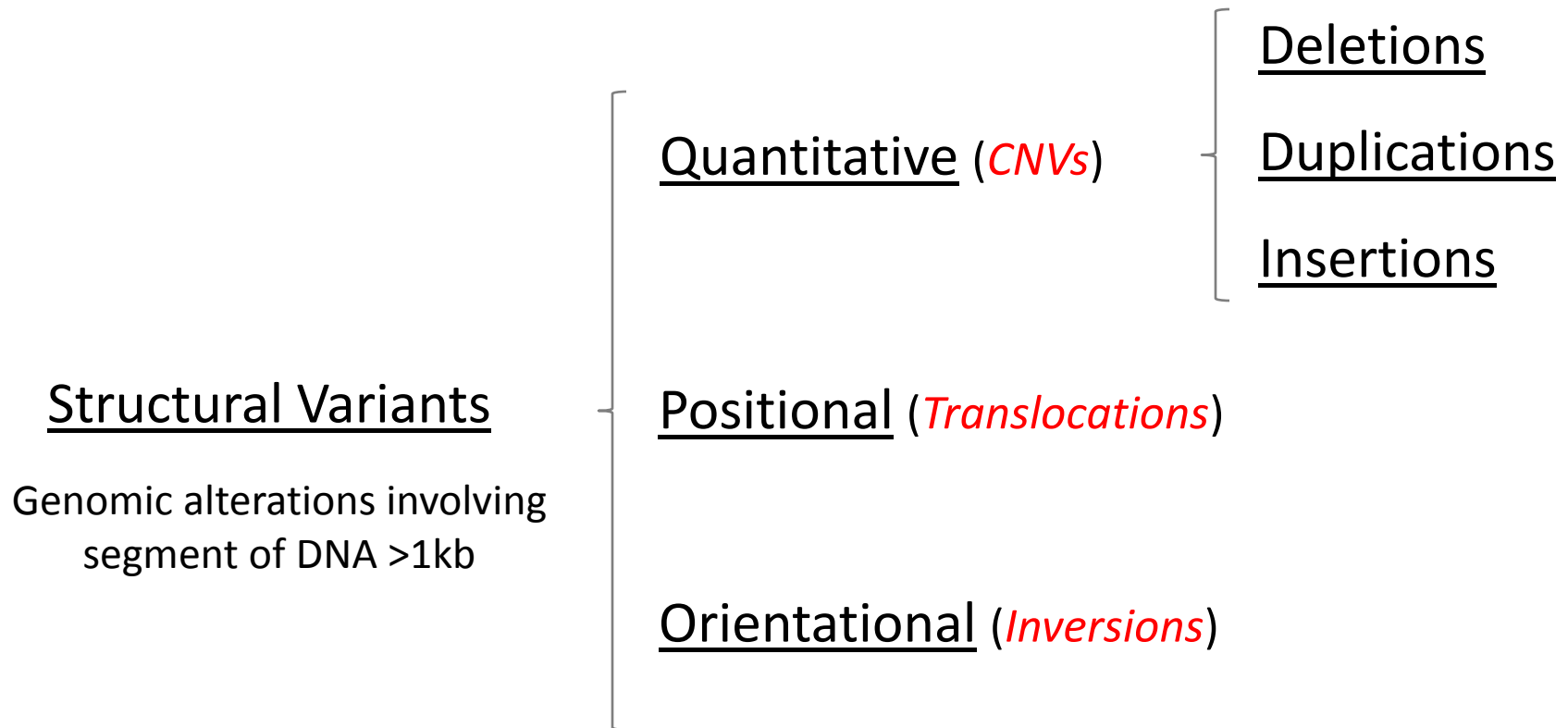
Outline

1. What is a Copy Number Variant (CNV)
2. Genome-wide detection of CNVs
3. Association analysis of CNVs
4. Online databases

1. What is a CNV?

What is a CNV?

1. Classes of structural variants



Copy Number Polymorphism (CNP) is a *CNV* that occurs in >1% population

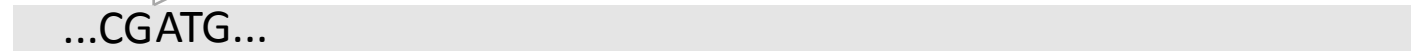
Duplication



1bp - Mb



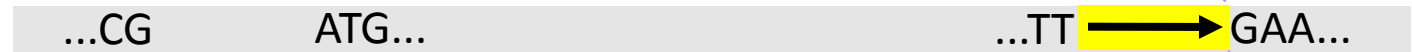
Deletion



Translocation



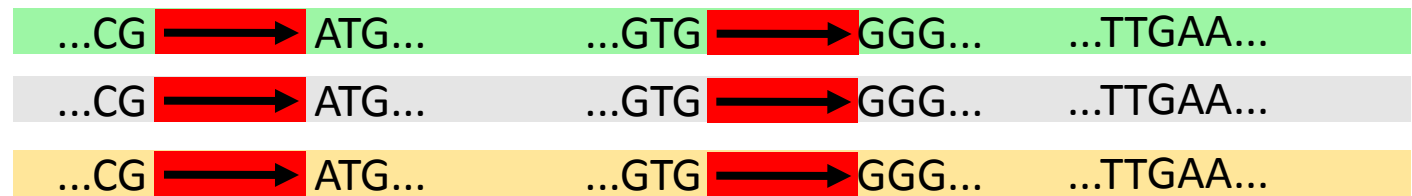
Insertion



Inversion



Segmental Duplication



With no CNV

Sequence variation

Single nucleotide

- Base change – substitution – point mutation
- Insertion-deletions (“indels”)
- SNPs – tagSNPs

Structural variation

2 bp to 1,000 bp

- Microsatellites, minisatellites
- Indels
- Inversions
- Di-, tri-, tetranucleotide repeats
- VNTRs

1 kb to submicroscopic

- Copy number variants (CNVs)
- Segmental duplications
- Inversions, translocations
- CNV regions (CNVRs)
- Microdeletions, microduplications

Microscopic to subchromosomal

- Segmental aneusomy
- Chromosomal deletions – losses
- Chromosomal insertions – gains
- Chromosomal inversions
- Intrachromosomal translocations
- Chromosomal abnormality
- Heteromorphisms
- Fragile sites

Whole chromosomal to whole genome

- Interchromosomal translocations
- Ring chromosomes, isochromosomes
- Marker chromosomes
- Aneuploidy
- Aneusomy

→ Term defined or discussed in **Box 1**

Molecular genetic detection

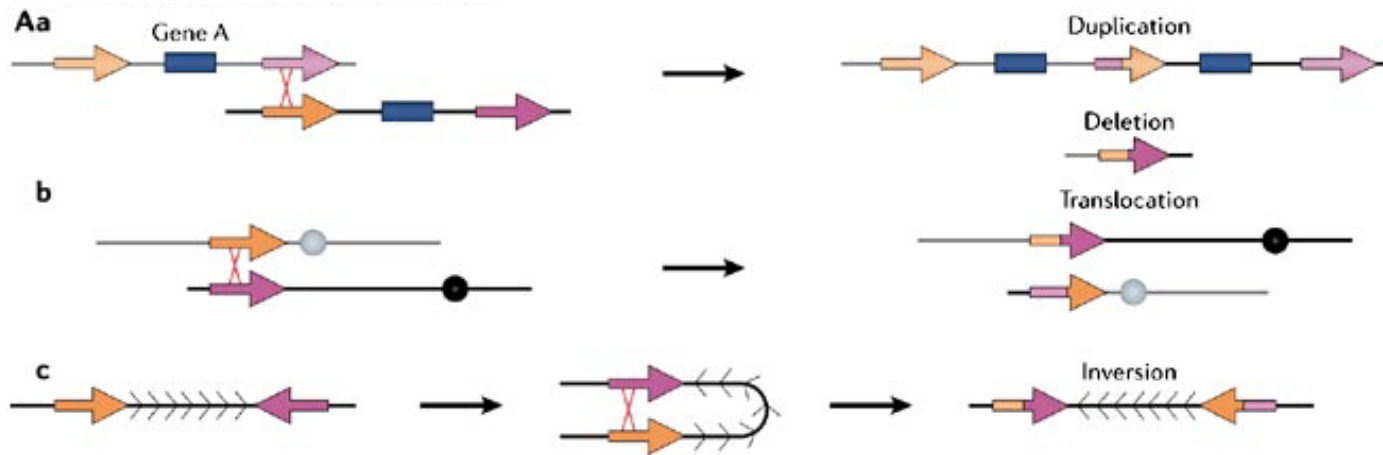


Cytogenetic detection

What is a CNV?

2. Origins of CNVs

(A) *Non-allelic homologous recombination*



(B) *Non-homologous end joining*

(C) *Tandem repeat sequences*

(D) *Retrotransposons*

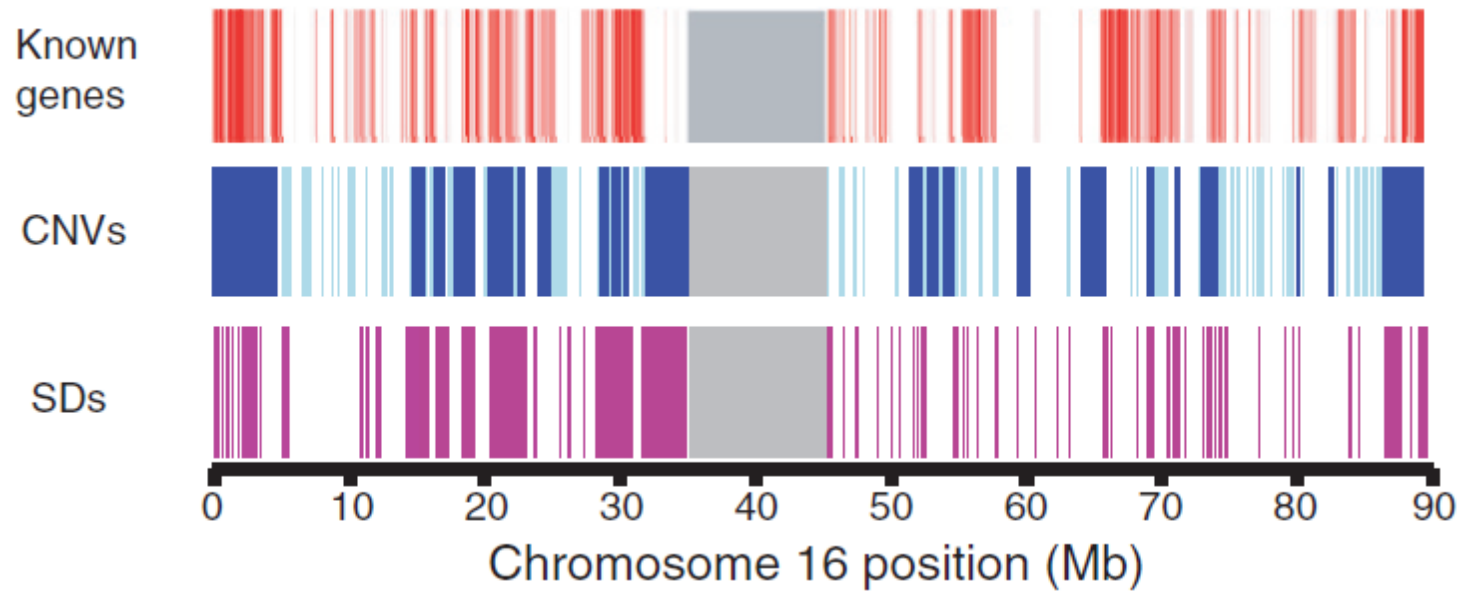
What is a CNV?

3. CNVs are abundant in the genome

Human vs Human	SNPs	CNVs
Base pairs	2.5 Mb	4 Mb
	1/1,200 bp	1/800
% genome	0.08%	0.12%

What is a CNV?

4. CNVs significantly overlap with known genes

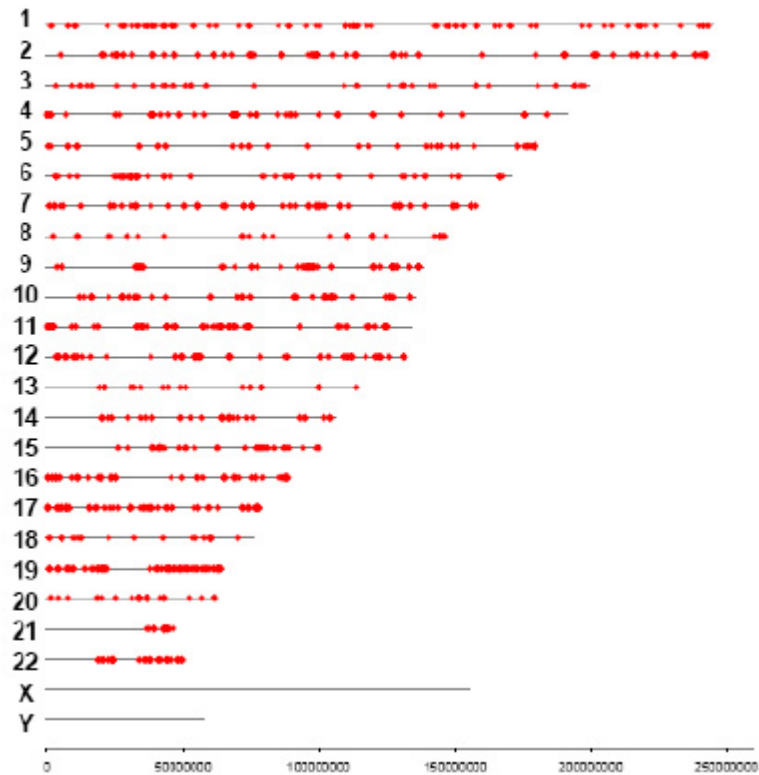


What is a CNV?

5. CNVs influence gene expression

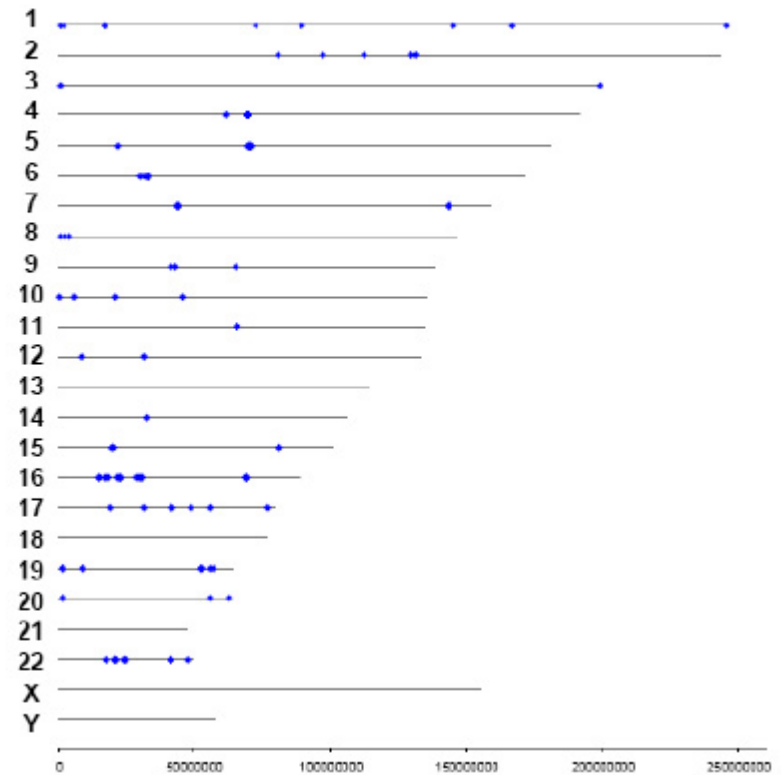
83.6%

A. SNP-expression association



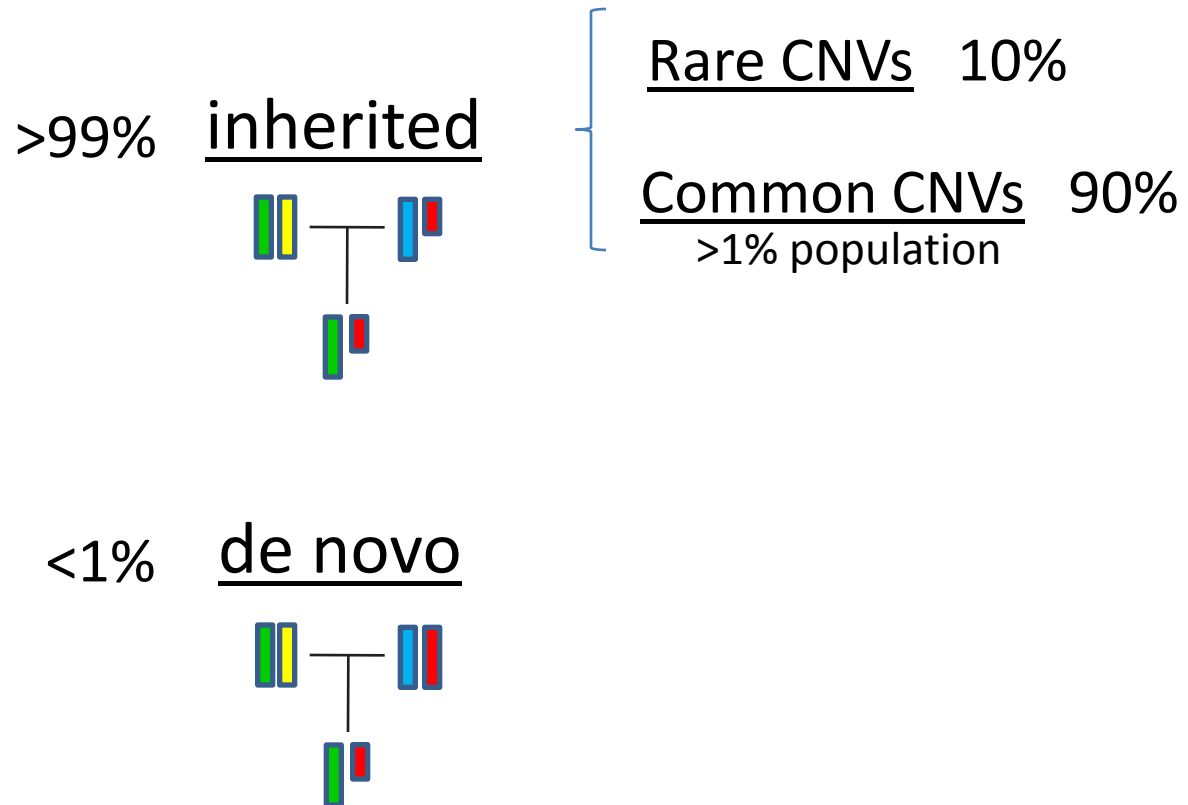
17.7%

B. CNV clone-expression association



What is a CNV?

6. In healthy individuals, most CNVs are inherited...



2. Detection of CNVs

Detection of CNVs

A. Using intensity data from whole-genome arrays

SNPs → Genotype *known common* variants

CNVs

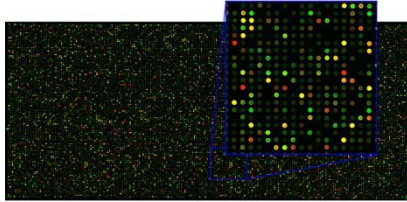
(A) Genotype *known common* variants

(B) *Identify and genotype new, potentially rarer* variants

Detection of CNVs

(A) Genotype *known common* CNVs using whole-genome arrays

Nimblegen



array-CGH, CNV only, test vs reference
custom or whole-genome (up to 2,1M probes)

Affy 6.0



>940,000 CNV non-polymorphic probes
High-density in ~5,600 CNV regions in DGV +
extended to whole-genome

Illumina 1M



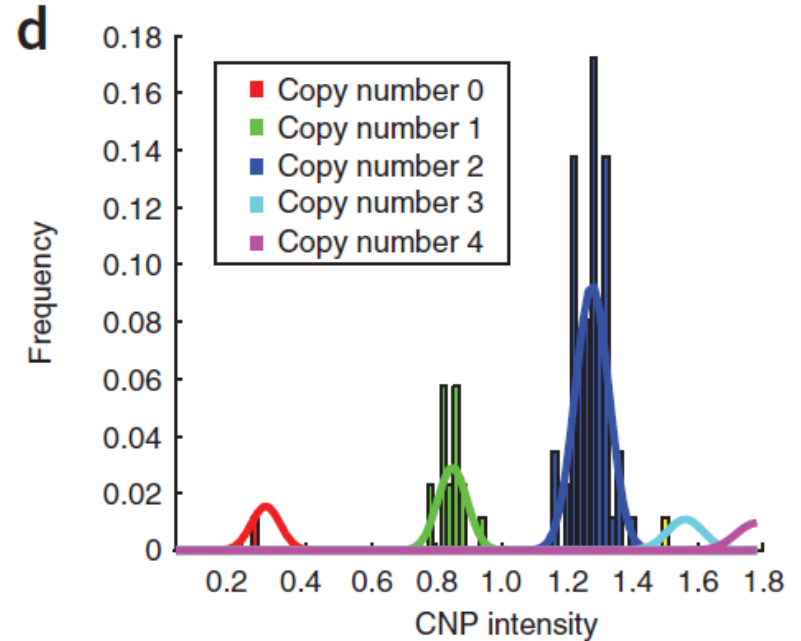
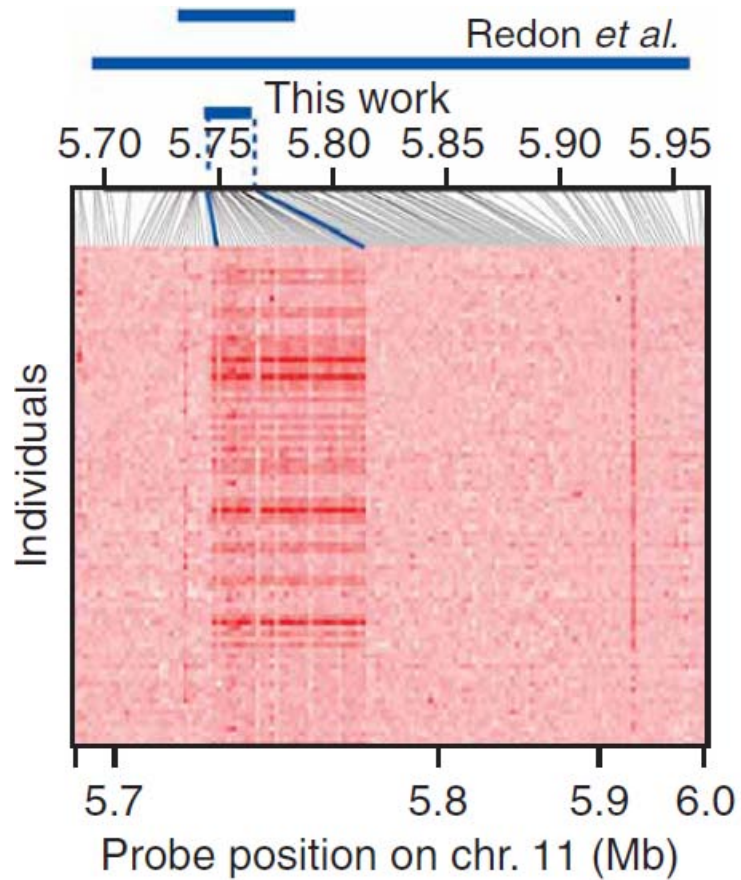
36,000 CNV non-polymorphic probes
covering ~4,000 CNV regions in DGV

Detection of CNVs



Ind	Genotype Mat/Pat	Copy number at S	Amount of DNA at S
1	S/S	2 <p>Two identical grey horizontal bars, each containing the sequence "...CG" followed by a red arrow pointing to "ATG...".</p>	<p>Two red arrows pointing upwards, indicating an increase in the amount of DNA at the S site.</p>

Detection of CNVs



Non-polymorphic probes

Detection of CNVs

(B) *Identify and genotype new, potentially rarer CNVs from whole-genome array data (CGH, Affymetrix/Illumina)*

Example: rs1006737 **A/G**

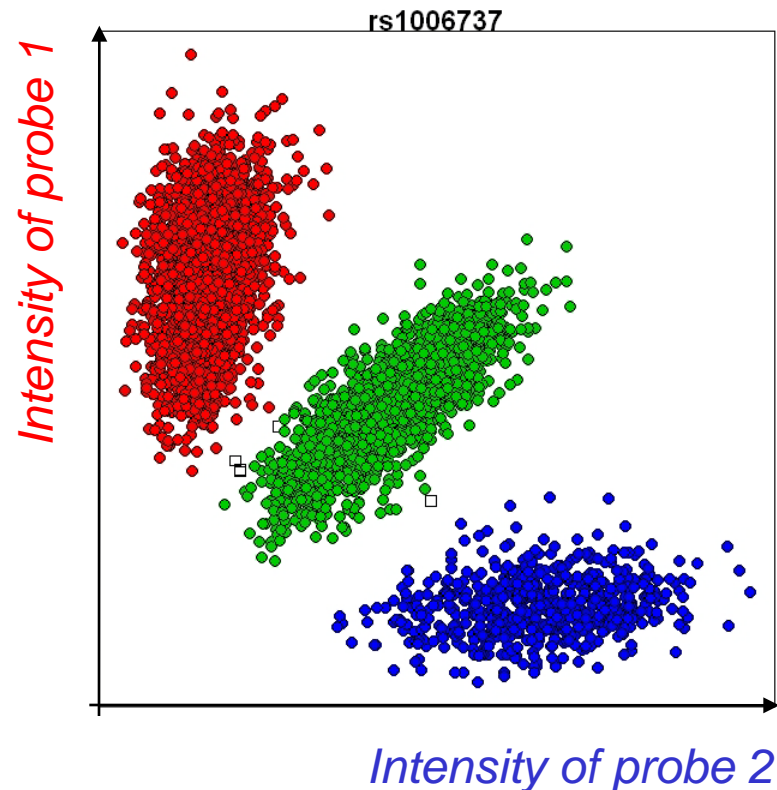
AA

AG

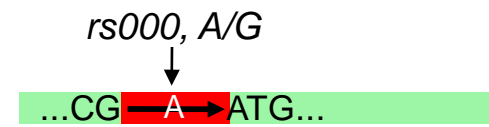
GG

... AGCCCGAAATGTTTTTCAGA... *probe 1*

... AGCCCGAAGTGTTTTTCAGA... *probe 2*



Detection of CNVs

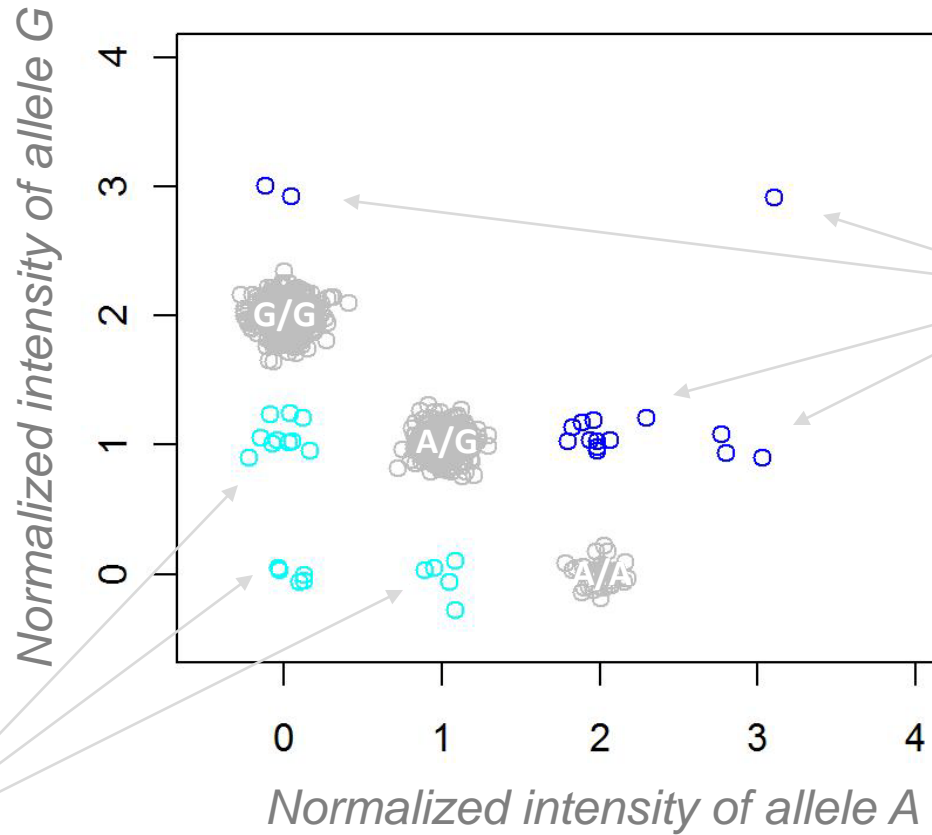


Ind	Genotype Mat/Pat	Pattern	Copy number for:		
			A	G	Total
1	A/G		1	1	2

Detection of CNVs

rs000, A/G

...CG **A** ATG...



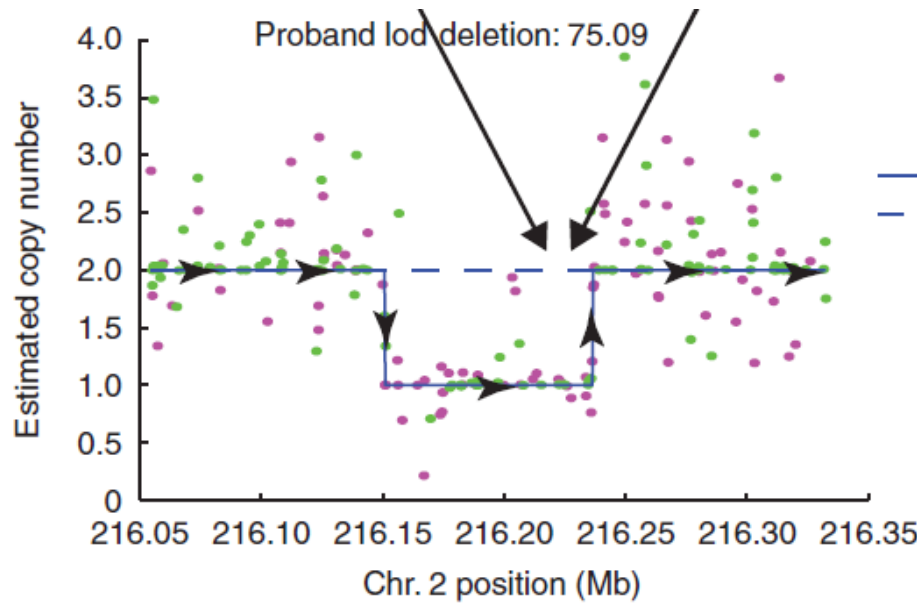
Individuals with duplication(s)
ie. total CN > 2

Individuals with deletion(s)

ie. total CN < 2

Polymorphic probe in CNV region

Detection of CNVs



Birdseye

Affy 5.0, 6.0

Korn et al 2008 Nat Genet 40: 1253

PennCNV

Affymetrix and Illumina

Wang et al 2007 Genome Res 17: 1665

Combine information across probes to identify new CNVs

For example...	Cases	Controls
100kb deletion chr. 2	10/5,000	1/5,000

Detecting CNVs through GWAS arrays is challenging...

Lots of confounders: DNA quality, concentration, source, batch effects, date effects.

Arrays have poor resolution for CNVs (>100kb).

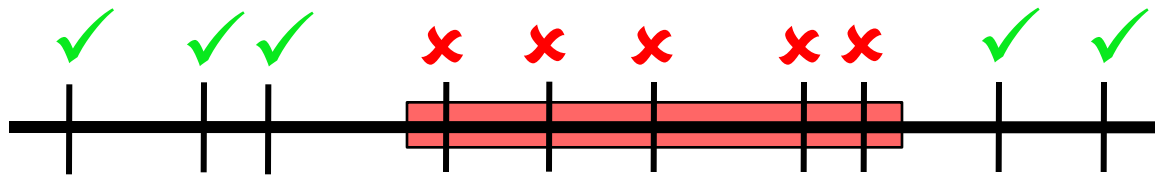
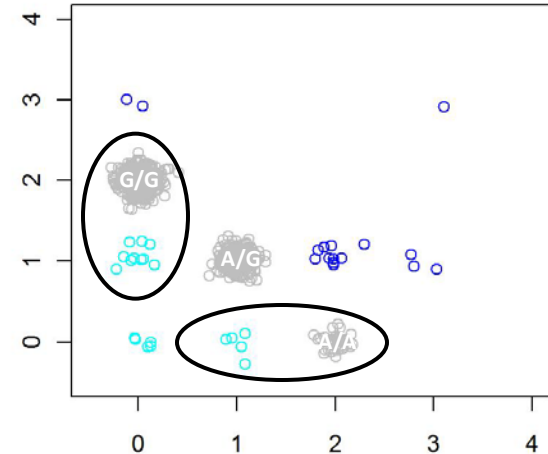
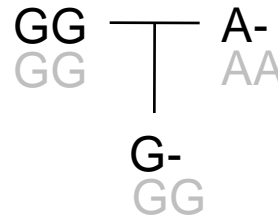
Genotype calling is computationally demanding, as it requires analysis of very large 'raw' cell files.

Genotype calling software often platform specific, not very user friendly.

Detection of CNVs

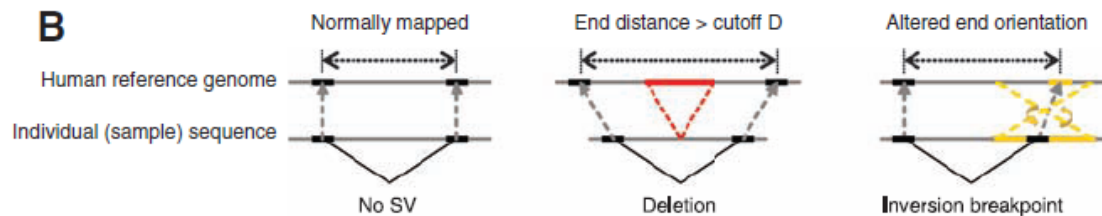
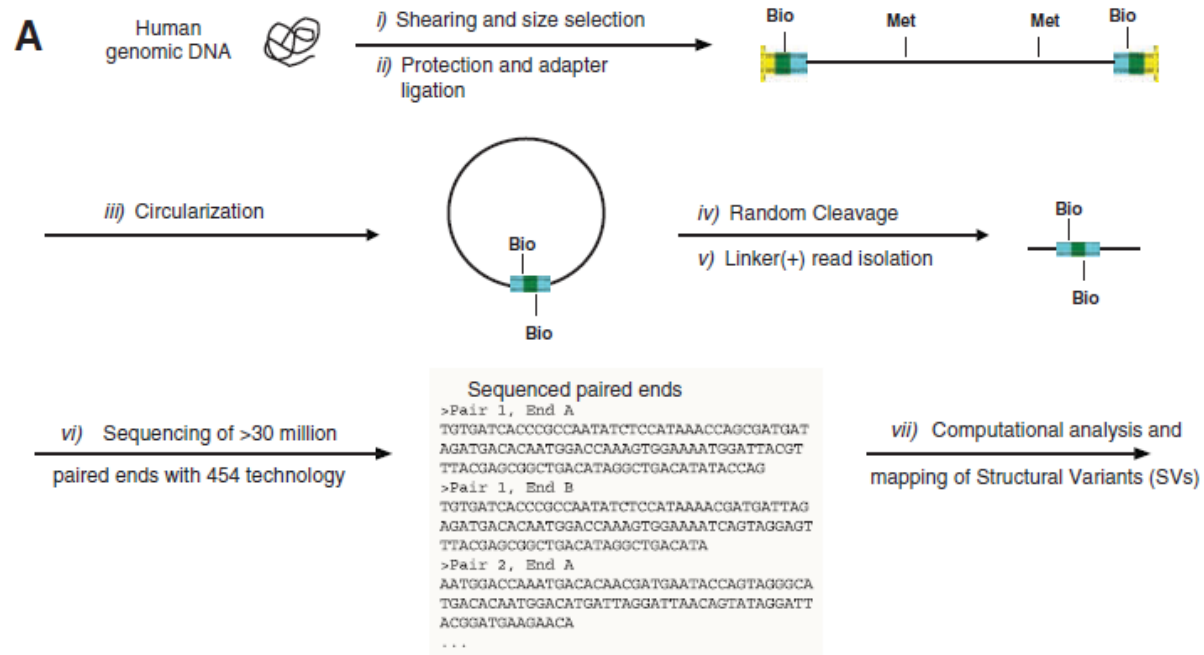
B. Identifying CNVs through genotyping errors

- ▶ Mendelian Inconsistencies
- ▶ Failure Hardy-Weinberg equilibrium



Detection of CNVs

C. Targeted or whole-genome sequencing



Summary so far...

CNVs are abundant, often overlap genes, can influence gene expression and most are inherited in healthy individuals

Known and new CNVs can be identified and genotyped in large-scale studies using whole-genome genotyping arrays, such as the 6.0 and 1M. Low resolution ($>100\text{Kb}$) & low signal/noise ratio.

More accurate CNV genotyping maps/arrays/algorithms expected in the next few years.

What are the particular strategies and challenges for association analysis of CNVs?

3. Association analysis of CNVs

Association analysis of CNVs

1. Some of the relevant questions

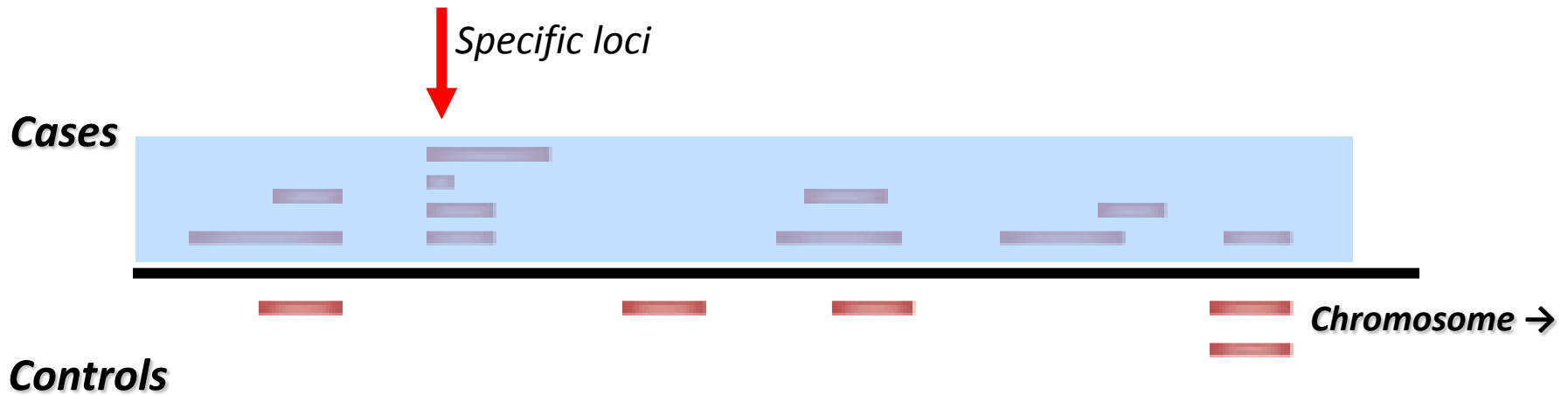
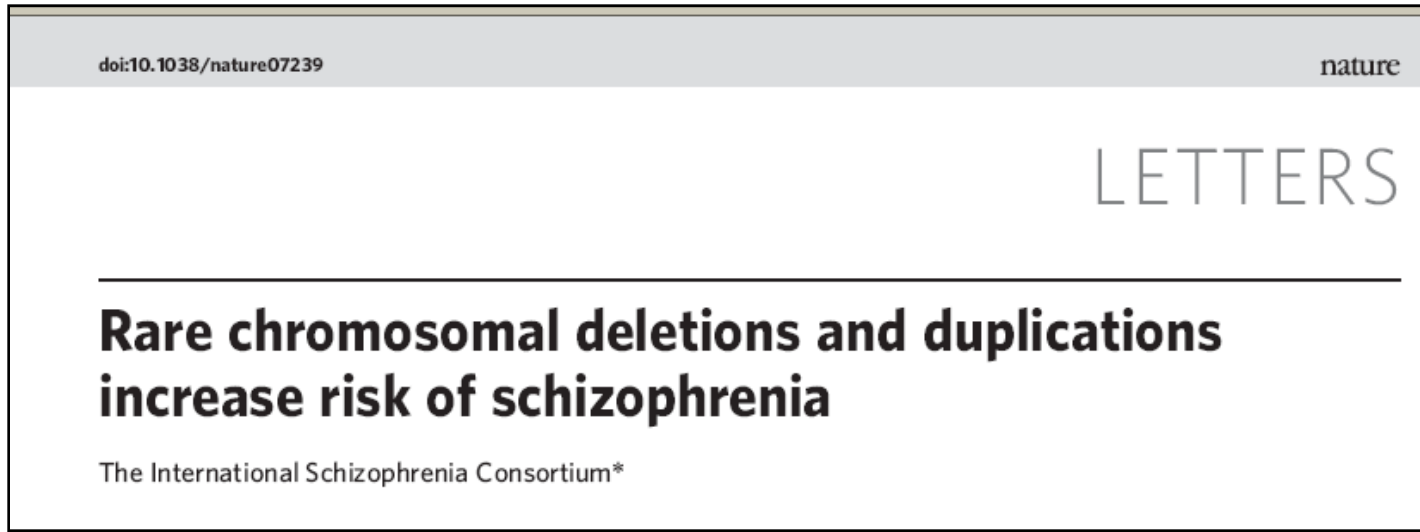
- (A) Are CNPs associated with variation in human traits or diseases?*
- (B) Can we identify rare CNVs associated with large increase in disease risk? Are these de novo or inherited in cases?*
- (C) When considering the whole-genome, do cases have more CNV events than controls, ie. increased burden?*
- (D) How to test SNPs in copy number regions?*
- (E) Are most CNVs tagged by SNPs in genotyping arrays?*

Example 1: Autism whole-genome CNV analysis

Sample	16p11	Cases	Controls	<i>P</i>	COPPER Birdseye CNAT
Discovery	Del (600kb)	5/1,441	3/4,234	1.1×10^{-4}	
[Affy 500K]	Dup	7/1,441	2/4,234		
Replication 1 (CHB)	Del	5/512	0/434	0.007	
[array-CGH]	Dup	4/512	0/434		
Replication 2 (deCODE)	Del	3/299	2/18,834	4.2×10^{-4}	
[Illumina]	Dup	0/299	5/18,834		

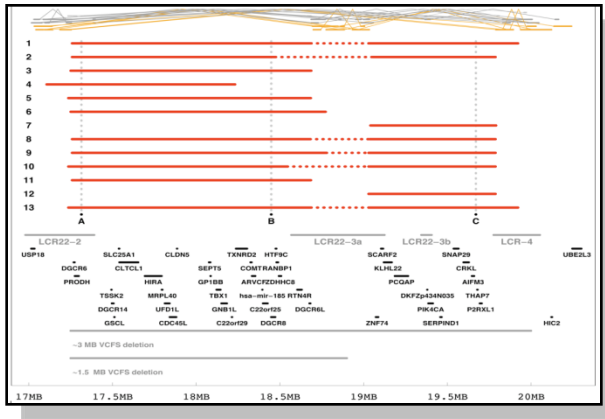
	<u>Deletion frequency Iceland</u>		<u>del</u>	<u>dup</u>
Autism	1%	inherited	2	6
Psychiatric disorder	0.1%	de novo	10	1
General population	0.01%	unknown	1	4

Example 2: SCZ whole-genome CNV analysis



Specific large (>500kb) rare deletions

22q11.2 (VCFS)



11 : 0

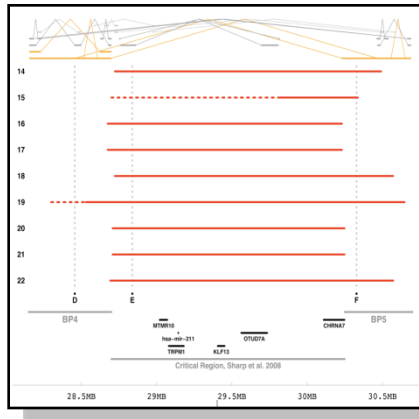
A “positive control”

*1:4000 live births
~30% develop psychosis
In ~0.6-2% SCZ patients*

*3Mb and 1.5Mb
variants*

*2 additional atypical
deletions observed*

15q13.3



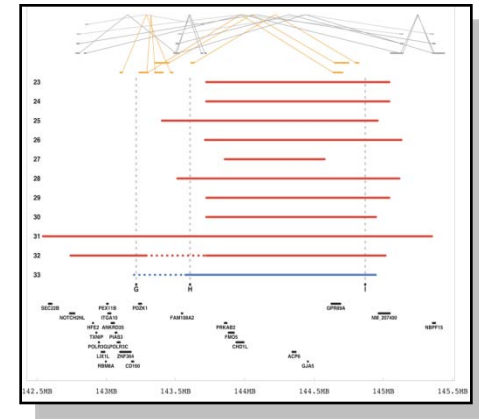
9 : 0

*CHRNA7, alpha 7
nicotinic acetylcholine
receptor*

*5 cases w/ impaired
cognition; 1 w/ epilepsy*

*Previously seen in mental
retardation with seizures*

1q21.1



10 : 1

*3 cases had cognitive
abnormalities; 1 with
epilepsy*

*Also seen in a patient
with MR and seizures
and two patients with
autism.*

Genome-wide burden of **rare** CNVs in SCZ

3,391 patients with SCZ, 3,181 controls
Filter for <1% MAF, >100kb
6,753 CNVs

Cases have greater rate of CNVs than controls

1.15-fold increase

$P = 3 \times 10^{-5}$

Cases have more genes intersected by CNVs than controls

1.14-fold increase

$P = 2 \times 10^{-6}$

**True for *singleton* events
(observed only once in dataset)**

1.45-fold increase

(~15% cases versus 11% controls)

$P = 5 \times 10^{-6}$

Rate of *genic* CNVs in cases v

1.18-fold increase

$P = 5 \times 10^{-6}$

CNVs in cases versus controls

1.09-fold increase

$P = 0.16$

Results invariant to obvious statistical controls

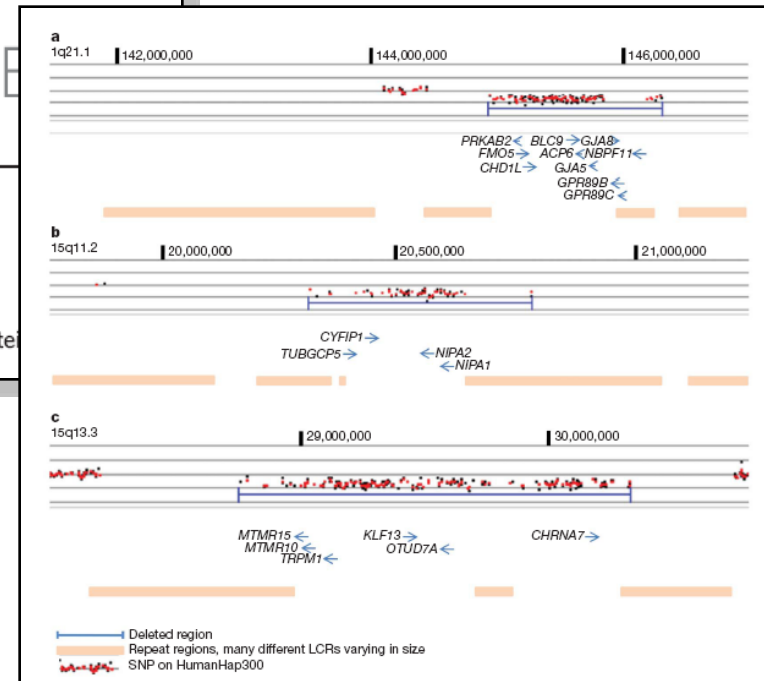
Array type, genotyping plate, sample collection site, mean probe intensity

LETTER

Large recurrent microdeletions associated with schizophrenia

Hreinn Stefansson^{1*}, Dan Rujescu^{2*}, Sven Cichon^{3,4*}, Olli P. H. Pietiläinen⁵, Andres Ingason¹, Stacy Steinberg¹, Ragnheidur Fossdal¹, Engilbert Sigurdsson⁶, Thordur Sigmundsson⁶, Jacobine E. Buizer-Voskamp⁷.





1q21.1 and 15q13.3 also identified by SGENE consortium

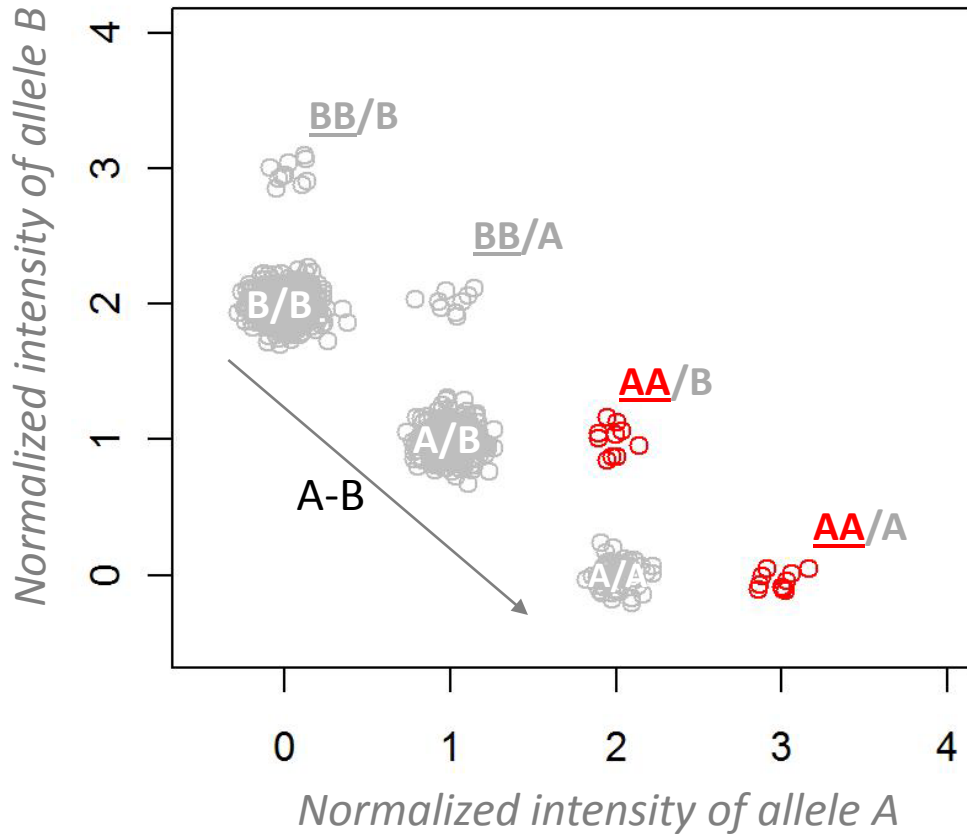


- Two other studies supporting a genome-wide increase in rare CNVs in schizophrenia
 - Walsh et al (2008) *Science*
 - 5% controls, 15% cases, 20% early onset cases
 - neurodevelopmental genes disrupted
 - Xu et al (2008) *Nature Genetics*
 - strong increased de novo rate in sporadic cases; but increased inherited rate also

Association analysis of CNVs

2. Testing SNPs in CNV regions

<p>Gene Z reference</p> 	Healthy	Healthy
<p>Gene Z with deletion</p> 	Disease	Healthy
<p>Gene Z with mutation</p> 	Disease	Healthy
<p>Gene Z with deletion and mutation</p> 	Disease	Disease
<p>Individual analysis of SNPs or CNVs</p>	✓	✗



$$y = \beta_1 \cdot \text{SNP} + \beta_2 \cdot \text{CNV}$$

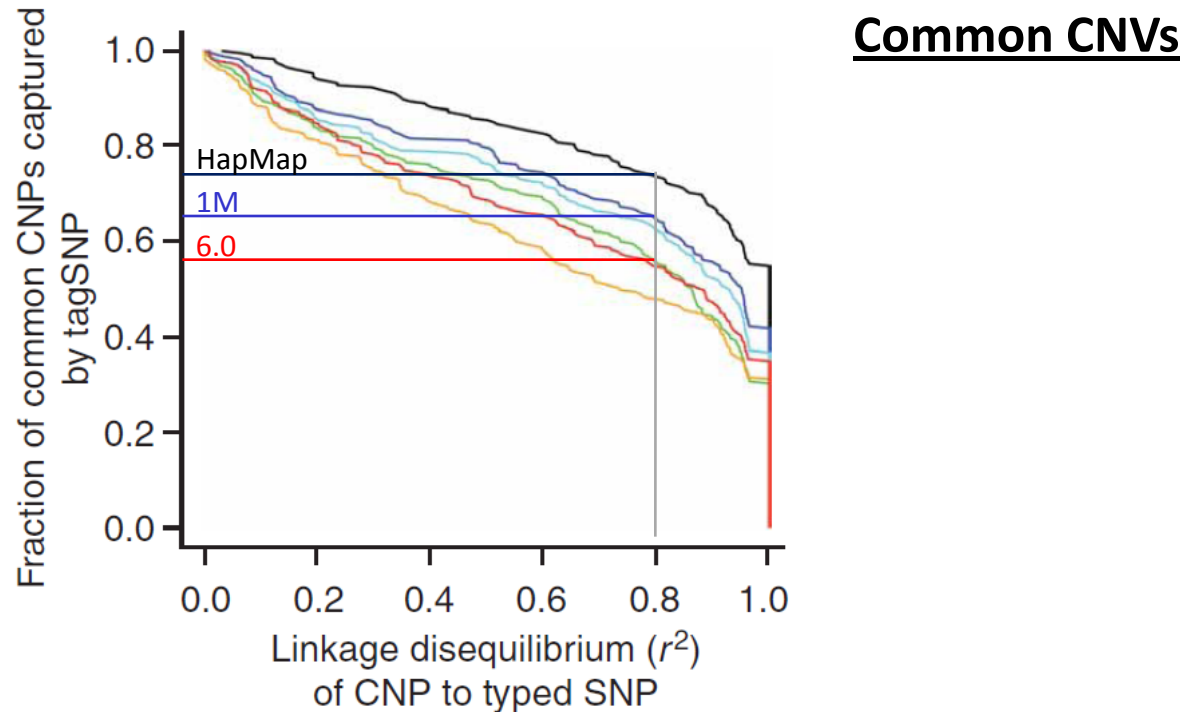


$$y = \beta_1 \cdot (A - B) + \beta_2 \cdot (A + B)$$

Allele-specific risk CNV

Association analysis of CNVs

3. Testing CNVs through the analysis of SNPs in LD



Coverage limited by lack of SNPs in CNV regions
(poor genotyping)

4. Online databases

Database of Genomic Variants

<http://projects.tcag.ca/variation/>

Database of Genomic Variants

A curated catalogue of structural variation in the human genome

Hosted by:
The Centre for
Applied
Genomics



[About The Project](#) | [Genome Browser](#) | [Download](#) | [Links](#) | [Data Submissions](#) | [Email us](#)

Please select genome assembly:

View Data by Chromosome

[1](#) [2](#) [3](#) [4](#) [5](#) [6](#) [7](#) [8](#) [9](#) [10](#) [11](#) [12](#) [13](#) [14](#) [15](#) [16](#) [17](#) [18](#) [19](#) [20](#) [21](#) [22](#) [X](#) [Y](#) [All](#)

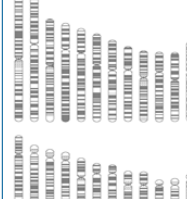
Keyword Search

Exact Match? Yes No
Examples: clone name, accession number, cytoband, gene

BLAT Search

Enter sequence in FASTA format here:

View Data by Genome



Summary Statistics

Total entries: **31615** (hg18)
CNVs: 19792
Inversions: 487
InDels (100bp-1Kb): 11336
Total CNV loci: 6225
Articles cited: 28
Last updated: Nov 10, 2008
Join our [mailing list](#)

Comprehensive summary of structural variation in the human genome. [Healthy control samples](#)

DECIPHER

<https://decipher.sanger.ac.uk/>

The screenshot shows the DECIPHER website homepage. At the top left is the Sanger Institute logo with the text 'welcome trust sanger institute'. A search bar is located at the top right. Below the logo is a navigation menu with items like 'DECIPHER', 'Database Entry Point', 'LOGIN', 'Patients & Projects', 'Syndromes', 'Join DECIPHER', 'DECIPHER Citations', 'Speed Test', 'Symposia', 'DECIPHER Converter', 'Information', 'Select', 'Documents', 'Resources', 'Contacts', 'Search by', 'Genomic Data', 'Website Search', 'People Search', 'Library Services', 'Site Map', and 'Feedback / Help'. The main content area features a 'Welcome to DECIPHER' message with a 'Guest Access' button. Below this is a 'Database of Chromosomal Imbalance and Phenotype in Humans using Ensembl Resources' section, followed by an 'About DECIPHER' section with a 'Background' subsection. The 'Background' section describes the database's purpose and resolution. There are also 'Challenges' and 'Solutions' sections. On the right side, there are three news letters: 'February News Letter' (dated 27th Feb 2009), 'January News Letter' (dated 23rd Jan 2009), and 'Genomic Disorders Conference' (dated 18th Dec 2008).

Database of submicroscopic chromosomal imbalances, from array-CGH data. Focuses on data from **patients** with developmental delay, learning disabilities or congenital anomalies.