

PLINK: a toolset for whole genome association analysis

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Challenges of whole-genome studies

- most obviously, multiple testing burden
- computationally-intensive methods break

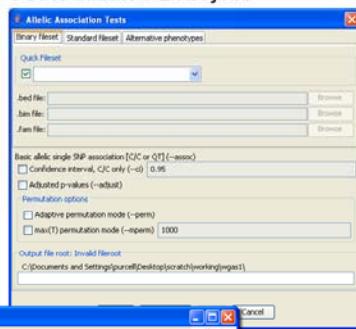
Opportunities

- simple methods can work well with ↑ data
- novel analyses permitted

PLINK

- introduce computational platform
- focus on basic data handling and testing
- “downstream analysis” (c.f. *Birdsuite*)

GUI to initiate PLINK jobs

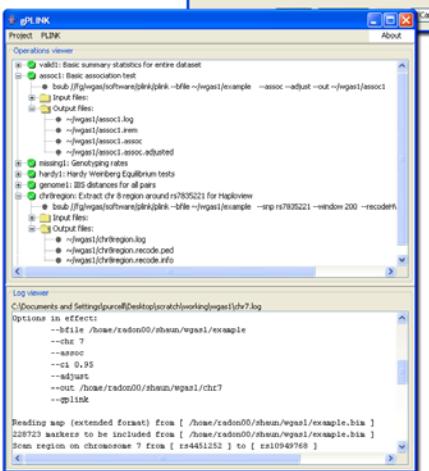


*Initiate PLINK jobs
locally or remotely*



Track PLINK jobs and results

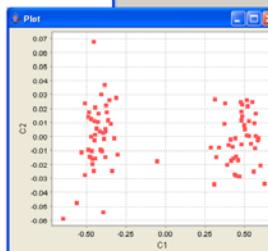
gPLINK



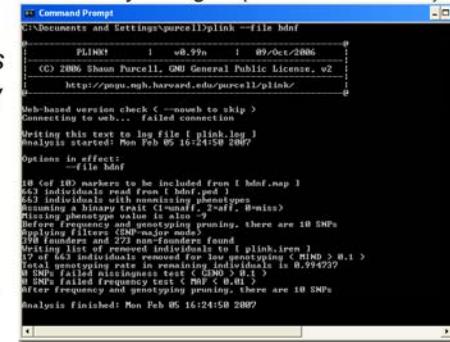
Job tracking interface

Integrate with Haploview

*Visualize PLINK
results (population
stratification)*

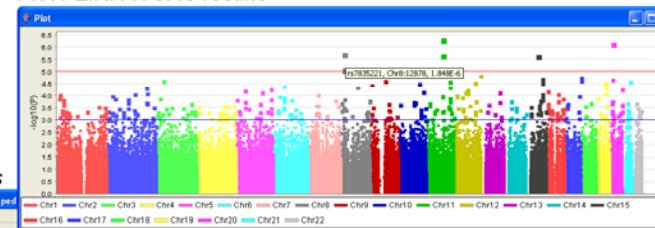


C/C++ analysis engine (can run standalone)



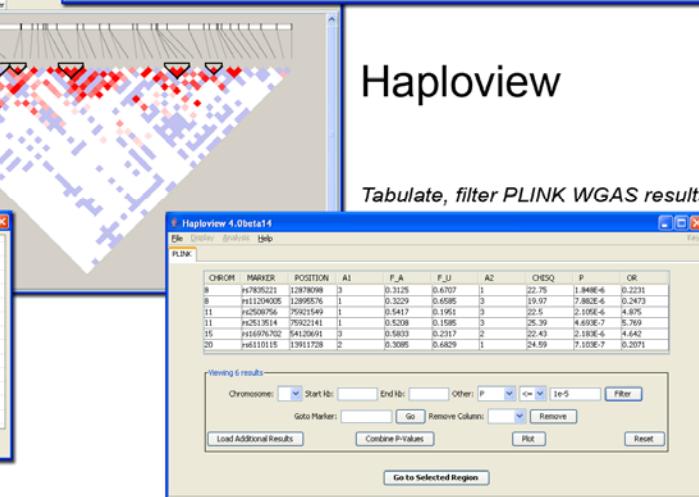
PLINK

Plot PLINK WGAS results



Haploview

Tabulate, filter PLINK WGAS results



- Data management
- Summary statistics
- Population stratification
- Association analysis
- Linkage disequilibrium and haplotype analysis
- Shared segment analysis
- Copy number analysis

Data management

SNPs →

P1	A A	A C	C G	T T	A A	T T
P2	A C	A A	C G	G T	A C	T T
P3	C C	A C	G G	T T	A A	T T
P4	C C	A A	G G	G T	A A	T T

← **People**

People →

S1	A A	A C	C C	C C
S2	A C	A A	A C	A A
S3	C G	C G	G G	G G
S4	T T	C G	T T	G T
S5	A A	G T	A A	A A
S6	T T	A C	T T	T T

← **SNPs**

P1	S1	A A
P1	S2	A C
P1	S3	C G
P2	S4	A C
P2	S5	C C

→ **Genotypes**

P1	S1	A	2	C	0
P1	S2	A	1	C	1
P1	S3	C	0	G	1
P2	S4	A	3	C	1
P2	S5	C	2	C	2

SNPs in CNPs

Compact binary format

```

010101001010101010101
1010011101010101010
1101110101001010101
1101001011101101010
1101010101010111010

```

	S1	S2	S3	S4
P1	0	1	2	1
P2	0	NA	0	2
P3	2	1	2	1
P4	0	1	2	0

Numeric coding

S1	A/A	P1	P2	P7	P8
S1	A/C	P3	P4		
S1	C/C	P5			
S2	G/T	P5			
S2	G/G	P1	P2	P3	P4

List by genotype

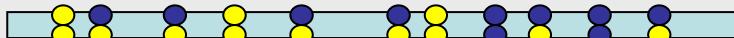
- Recode dataset (A,C,G,T → 1,2)
- Reorder, reformat dataset
- Flip DNA strand
- Extract/remove individuals/SNPs
- Swap in new phenotypes, covariates
- Filter on covariates
- Merge 2 or more filesets

Summary statistics

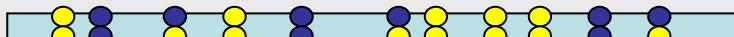
- Filters and reports for standard metrics
 - Genotyping rate
 - Allele, genotype, haplotype frequencies
 - Hardy-Weinberg
 - Mendel errors
- Tests of non-random missingness
 - by phenotype and by (unobserved) genotype
- Individual homozygosity estimates
- Check/impute sex based on X chromosome
- LD-based detection of strand flips
 - A/T and C/G SNPs potentially ambiguous
- Automated search for plate effects
 - w/ subsequent masking of specific SNP/individual genotypes

Pairwise allele-sharing metric

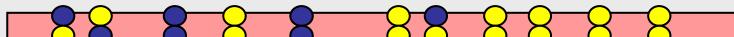
Reference



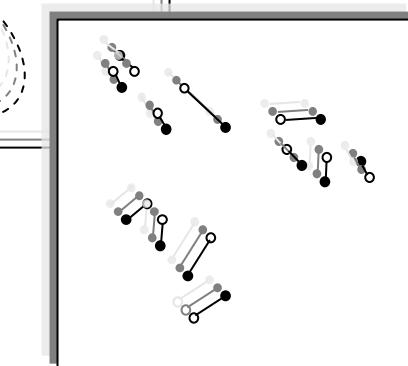
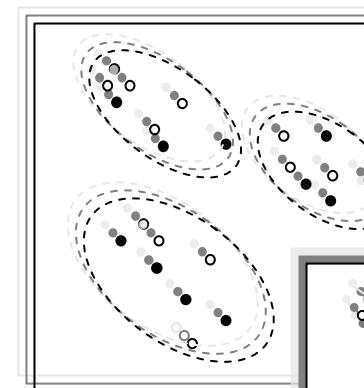
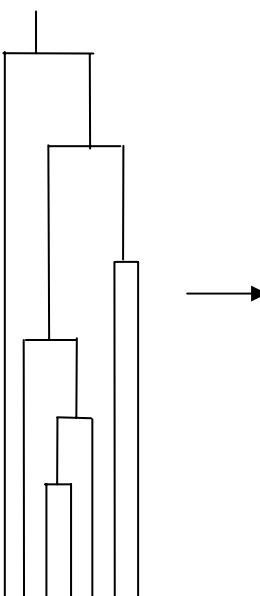
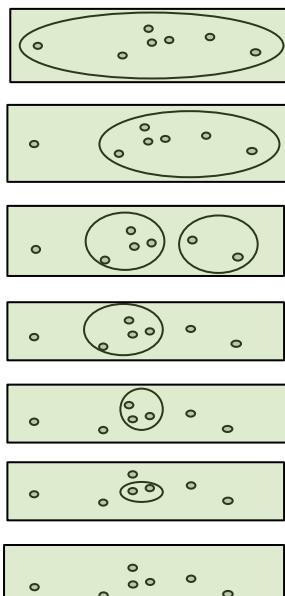
Same population



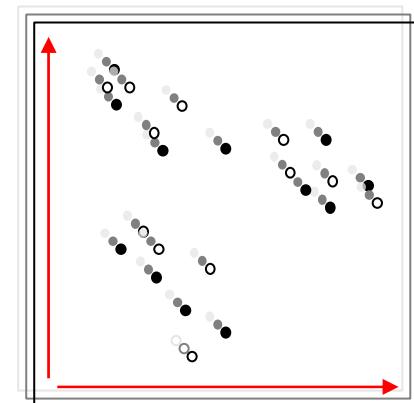
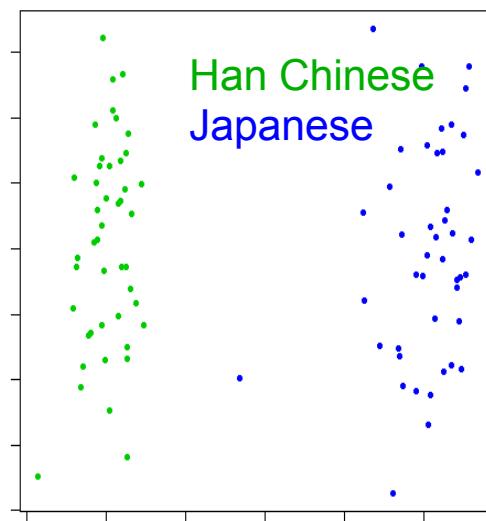
Different population



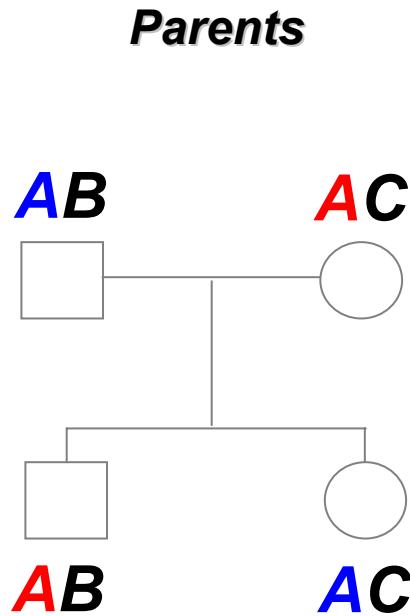
Hierarchical clustering



Multidimensional scaling/PCA



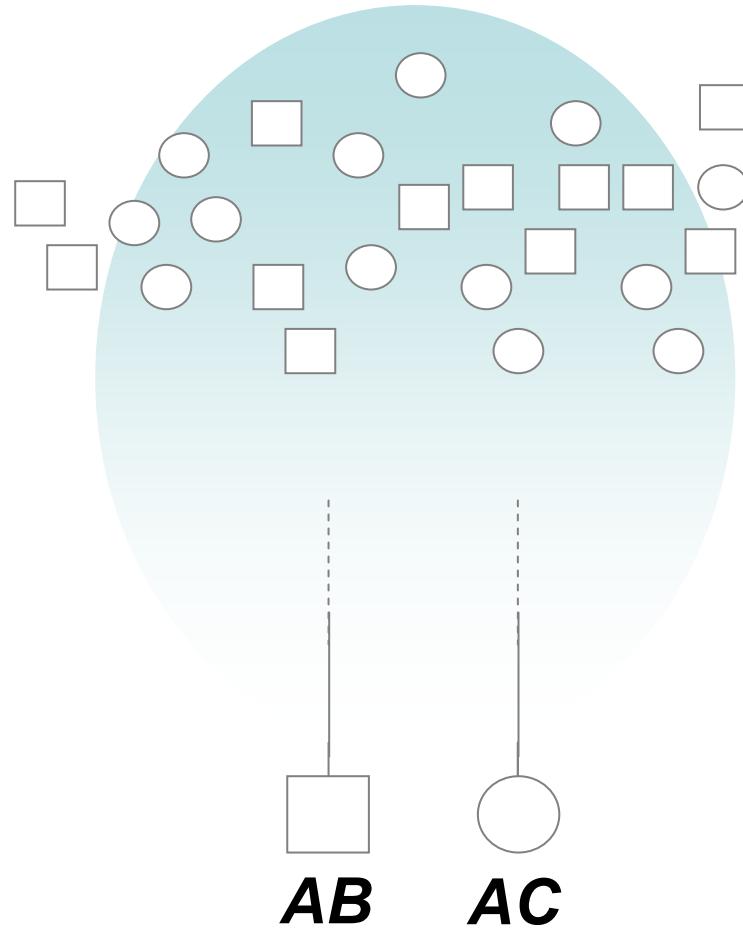
Estimation of IBD sharing (relatedness)



$$IBS = 1$$

$$IBD = 0$$

Most recent common ancestor from homogeneous random mating population



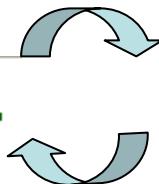
Association analysis

- Population-based
 - Allelic, trend, genotypic, Fisher's exact
 - Stratified tests (Cochran-Mantel-Haenszel, Breslow-Day)
 - Linear & logistic regression models
 - multiple covariates, interactions, joint tests, etc
- Family-based
 - Disease traits: TDT / sib-TDT
 - Continuous traits: QFAM (between/within model, QTDT)
- Permutation procedures
 - “adaptive”, max(T), gene-dropping, between/within, rank-based, within-cluster
- Multilocus tests
 - Haplotype estimation, set-based tests, Hotelling's T^2 , epistasis

R plugin support

SNP data

plink...



e.g.
Survival analysis
Multinomial regression
Log-linear models

Output

Runs of homozygosity, extended haplotype sharing



Chromosome →

Disease
locus

Simple SNP simulation functions



SIMULATE

P1	A	A	A	C	C	G	T	T
P2	A	C	A	A	C	G	G	T
P3	C	C	A	C	G	G	T	T
P4	C	C	A	A	G	G	G	T

```
plink --lookup rs222162
```

```
PLINK-SNP (WGAS SNP annotation courtesy of Patrick Sullivan)  
Connecting to web...
```

SNP ID	:	rs222162
Affy 5.0	:	no
Affy 6.0	:	no
Perlegen ID	:	43887
Perlgen 600	:	yes
Illumina 650	:	yes
Non-syn SNP	:	no
SNP Error	:	no
Chromosome	:	21
Strand	:	+
Position (bp)	:	26755483
Pseudo-autosomal region?	:	N/A
NCBI reference allele	:	G
Human alleles	:	A/G
Chimp allele	:	A
HapMap CEU MAF	:	0.075
HapMap ASI MAF	:	0
HapMap YRI MAF	:	0.5
In gene transcript	:	AK125338
Nearby Genes(KB distance)	:	A4(-290) CYYR1 (4) ATS1 (374) ATS5 (460)
Segmental duplication?	:	no
Conservation >95% pctile?	:	no
miRNA target? (TargetScan)	:	no
Regulatory potential?	:	yes
Promotor region? (Stanford)	:	no
Transfactor binding site	:	no
Enhancer?	:	no
Exon?	:	no
Consensus splice site?	:	no
5' UTR?	:	no
3' UTR?	:	no

PLINK: Whole genome data analysis toolset - Mozilla Firefox

File Edit View History Bookmarks Tools Help

http://pngu.mgh.harvard.edu/~purcell/plink/ plink

SquirrelMail 1.4.8 Getting Started Latest Headlines

plink...

Latest PLINK release is v1.01 (28-Jan-2008)

Whole genome association analysis toolset

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1. Introduction

2. Basic information

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- ◆ [Reporting problems](#)
- ◆ [What's new?](#)

3. Download and general notes

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- ◆ [Development code](#)
- ◆ [General notes](#)
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- ◆ [Unix/Linux notes](#)
- ◆ [Compilation](#)
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- ◆ [Under development](#)

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- ◆ [PED files](#)
- ◆ [MAP files](#)
- ◆ [Transposed filesets](#)
- ◆ [Long-format filesets](#)
- ◆ [Binary PED files](#)
- ◆ [Alternate phenotypes](#)
- ◆ [Covariate files](#)

PLINK is a free, open-source whole genome association analysis toolset, designed to perform a range of basic, large-scale analyses in a computationally efficient manner.

The focus of **PLINK** is purely on *analysis* of genotype/phenotype data, so there is no support for steps prior to this (e.g. study design and planning, generating genotype calls from raw data). Through integration with [gPLINK](#) and [Haploview](#), there is some support for the subsequent visualization, annotation and storage of results.

PLINK (one syllable) is being developed by Shaun Purcell at the Center for Human Genetic Research (CHGR), Massachusetts General Hospital (MGH), and the [Broad Institute](#) of Harvard & MIT, with the support of others.

Quick links

[PLINK tutorial](#)
[gPLINK](#)
[Join e-mail list](#)
[Resources](#)
[FAQs](#)
[Citing PLINK](#)
[Bugs, questions?](#)

Data management

- ◆ Read data in a variety of formats
- ◆ Recode and reorder files
- ◆ Merge two or more files
- ◆ Extracts subsets (SNPs or individuals)
- ◆ Flip strand of SNPs
- ◆ Compress data in a binary file format

Summary statistics for quality control

Documentation also available as PDF (>200 pages)

A simulated WGAS dataset



Summary statistics and quality control



Whole genome SNP-based association



Whole genome haplotype-based association



Assessment of population stratification



Further exploration of ‘hits’



Visualization and follow-up using Haploview

Acknowledgements

- **PLINK development**
 - Kathe Todd-Brown
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 - Ben Neale
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 - Itsik Pe'er
 - Ben Voight
 - David Altshuler
 - Pamela Sklar
 - Mark Daly

PLINK practical 1

Data cleaning and
association testing

- In this practical we will analyse a simulated dataset using PLINK
- 15 single nucleotide polymorphisms, in a candidate gene region spanning ~30kb
- Case/control design: 1000 cases and 1000 controls
- Specifically, we will:
 - examine the format of the raw data (PED and MAP files)
 - perform an initial association analysis for each SNP
 - perform basic QC steps, including tests for HWE and looking at genotyping rate statistics
 - repeat the association analysis
 - consider genotypic as well as allelic tests
 - perform sex-specific tests
 - perform conditional and haplotypic tests

Preliminary association analysis (allelic)

```
./plink --file mygene  
--assoc
```

Output file plink.assoc

CHR	SNP	BP	A1	F_A	F_U	A2	CHISQ	P	OR
1	rs00001	0	G	0.13	0.1524	C	4.015	0.04509	0.8314
1	rs00002	2013	C	0.1489	0.1459	A	0.07039	0.7908	1.024
1	rs00003	4367	A	0.4612	0.5225	T	11.92	0.0005542	0.7825
1	rs00004	6473	G	0.4286	0.4279	C	0.002133	0.9632	1.003
1	rs00005	8887	C	0.454	0.4636	T	0.3656	0.5454	0.9619
1	rs00006	11054	T	0.1725	0.1619	A	0.7824	0.3764	1.079
1	rs00007	13413	G	0.03846	0.04239	A	0.3887	0.533	0.9036
1	rs00008	15820	T	0.139	0.1014	G	13.14	0.0002883	1.43
1	rs00009	18125	T	0.2391	0.2024	C	7.681	0.005582	1.238
1	rs00010	20253	A	0.423	0.4117	C	0.5109	0.4748	1.048
1	rs00011	22633	C	0.1925	0.1926	G	8.647e-05	0.9926	0.9992
1	rs00012	24739	C	0.152	0.1326	T	2.995	0.08353	1.172
1	rs00013	26762	G	0.2829	0.5571	A	302.3	1.041e-67	0.3136
1	rs00014	28833	A	0.1985	0.2161	G	1.856	0.1731	0.8982
1	rs00015	30974	A	0.2071	0.225	C	1.849	0.1739	0.8996

Allele frequencies

```
./plink --file mygene  
--freq
```

Output file plink.frq

CHR	SNP	A1	A2	MAF	NCHROBS
1	rs00001	G	C	0.1412	3902
1	rs00002	C	A	0.1474	3914
1	rs00003	A	T	0.486	3296
1	rs00004	G	C	0.4283	3918
1	rs00005	C	T	0.4588	3908
1	rs00006	T	A	0.1672	3924
1	rs00007	G	A	0.04043	3908
1	rs00008	T	G	0.1202	3936
1	rs00009	T	C	0.2208	3936
1	rs00010	A	C	0.4174	3896
1	rs00011	C	G	0.1925	3932
1	rs00012	C	T	0.1423	3892
1	rs00013	G	A	0.4201	3918
1	rs00014	A	G	0.2073	3922
1	rs00015	A	C	0.2161	3906

Genotyping rate per individual and per marker

```
./plink --file mygene  
--missing
```

*Per-individual genotyping/missing
rate, plink.imiss*

FID	IID	MISS_PHENO	N_MISS	N_GENO	F_MISS
per0	per0	N	0	15	0
per1	per1	N	0	15	0
per2	per2	N	0	15	0
per3	per3	N	0	15	0
per4	per4	N	0	15	0
per5	per5	N	1	15	0.06667
per6	per6	N	0	15	0
per7	per7	N	2	15	0.1333
...	...				

*Per-marker (locus) genotyping/
missing rate, plink.lmiss*

CHR	SNP	N_MISS	N_GENO	F_MISS
1	rs00001	49	2000	0.0245
1	rs00002	43	2000	0.0215
1	rs00003	352	2000	0.176
1	rs00004	41	2000	0.0205
1	rs00005	46	2000	0.023
1	rs00006	38	2000	0.019
1	rs00007	46	2000	0.023
1	rs00008	32	2000	0.016
1	rs00009	32	2000	0.016
1	rs00010	52	2000	0.026
1	rs00011	34	2000	0.017
1	rs00012	54	2000	0.027
1	rs00013	41	2000	0.0205
1	rs00014	39	2000	0.0195
1	rs00015	47	2000	0.0235

Check for Hardy-Weinberg disequilibrium

```
./plink --file mygene  
--hardy
```

Output file plink.frq

CHR	SNP	TEST	A1	A2	GENO	O(HET)	E(HET)	P
1	rs00001	ALL	G	C	44/463/1444	0.2373	0.2425	0.3501
1	rs00001	AFF	G	C	17/219/737	0.2251	0.2262	0.8871
1	rs00001	UNAFF	G	C	27/244/707	0.2495	0.2583	0.2674
1	rs00002	ALL	C	A	34/509/1414	0.2601	0.2514	0.1495
1	rs00002	AFF	C	A	17/257/703	0.2631	0.2535	0.3114
1	rs00002	UNAFF	C	A	17/252/711	0.2571	0.2493	0.3707
1	rs00003	ALL	A	T	415/772/461	0.4684	0.4996	0.0119
1	rs00003	AFF	A	T	215/474/291	0.4837	0.497	0.4038
1	rs00003	UNAFF	A	T	200/298/170	0.4461	0.499	0.006616
1	rs00004	ALL	G	C	363/952/644	0.486	0.4897	0.7467
1	rs00004	AFF	G	C	178/485/318	0.4944	0.4898	0.7945
1	rs00004	UNAFF	G	C	185/467/326	0.4775	0.4896	0.4339
...	...							
1	rs00013	ALL	G	A	567/512/880	0.2614	0.4872	5.262e-96
1	rs00013	AFF	G	A	277/0/702	0	0.4058	7.409e-254
1	rs00013	UNAFF	G	A	290/512/178	0.5224	0.4935	0.07019

Check for differential genotyping rate (case vs. control)

```
./plink --file mygene  
--test-missing
```

Output file plink.missing

CHR	SNP	F_MISS_A	F_MISS_U	P
1	rs00001	0.027	0.022	0.5633
1	rs00002	0.023	0.02	0.7582
1	rs00003	0.02	0.332	6.43e-87
1	rs00004	0.019	0.022	0.7527
1	rs00005	0.022	0.024	0.8816
1	rs00006	0.023	0.015	0.2513
1	rs00007	0.025	0.021	0.655
1	rs00008	0.018	0.014	0.5936
1	rs00009	0.015	0.017	0.8589
1	rs00010	0.026	0.026	1
1	rs00011	0.018	0.016	0.863
1	rs00012	0.023	0.031	0.3343
1	rs00013	0.021	0.02	1
1	rs00014	0.02	0.019	1
1	rs00015	0.027	0.02	0.376

Remove bad SNPs

```
echo "rs00003" > bad.snps
echo "rs00013" >> bad.snps

./plink --file mygene
    --exclude bad.snps
    --recode
    --out cleaned
```

Equivalent command, using filters

```
./plink --file mygene
    --hwe 1e-3
    --hwe-all
    --geno 0.1
    --recode
    --out cleaned
```

Normally, HWE filters on controls; the --hwe-all flag implies all individuals

Re-run association analysis (allelic)

```
./plink --file cleaned  
--assoc
```

Output file plink.assoc

CHR	SNP	BP	A1	F_A	F_U	A2	CHISQ	P	OR
1	rs00001	0	G	0.13	0.1524	C	4.015	0.04509	0.8314
1	rs00002	2013	C	0.1489	0.1459	A	0.07039	0.7908	1.024
1	rs00004	6473	G	0.4286	0.4279	C	0.002133	0.9632	1.003
1	rs00005	8887	C	0.454	0.4636	T	0.3656	0.5454	0.9619
1	rs00006	11054	T	0.1725	0.1619	A	0.7824	0.3764	1.079
1	rs00007	13413	G	0.03846	0.04239	A	0.3887	0.533	0.9036
1	rs00008	15820	T	0.139	0.1014	G	13.14	0.0002883	1.43
1	rs00009	18125	T	0.2391	0.2024	C	7.681	0.005582	1.238
1	rs00010	20253	A	0.423	0.4117	C	0.5109	0.4748	1.048
1	rs00011	22633	C	0.1925	0.1926	G	8.647e-05	0.9926	0.9992
1	rs00012	24739	C	0.152	0.1326	T	2.995	0.08353	1.172
1	rs00014	28833	A	0.1985	0.2161	G	1.856	0.1731	0.8982
1	rs00015	30974	A	0.2071	0.225	C	1.849	0.1739	0.8996

Corrections for multiple testing

```
./plink --file cleaned  
    --assoc  
    --adjust  
    --mperm 10000
```

Output file plink.assoc.adjust: rs00008 is significant after Bonferroni correction

CHR	SNP	UNADJ	GC	BONF	HOLM	SIDAK_SS	SIDAK_SD	FDR_BH	FDR_BY
1	rs00008	0.0002883	0.005643	0.003748	0.003748	0.003742	0.003742	0.003748	0.01192
1	rs00009	0.005582	0.03437	0.07256	0.06698	0.07018	0.06496	0.03628	0.1154
1	rs00001	0.04509	0.1261	0.5862	0.496	0.4511	0.398	0.1954	0.6214
1	rs00012	0.08353	0.1864		0.8353	0.6782	0.582	0.2715	0.8633
1	rs00014	0.1731	0.2983	1	1	0.9155	0.8192	0.3768	1
1	rs00015	0.1739	0.2992	1	1	0.9166	0.8192	0.3768	1
1	rs00006	0.3764	0.4995	1	1	0.9978	0.9633	0.699	1
1	rs00010	0.4748	0.5853	1	1	0.9998	0.979	0.709	1
1	rs00007	0.533	0.6341	1	1	0.9999	0.979	0.709	1
1	rs00005	0.5454	0.6444	1	1	1	0.979	0.709	1
1	rs00002	0.7908	0.8395	1	1	1	0.9908	0.9345	1
1	rs00004	0.9632	0.9719	1	1	1	0.9986	0.9926	1
1	rs00011	0.9926	0.9943	1	1	1	0.9986	0.9926	1

Corrections for multiple testing

```
./plink --file cleaned  
    --assoc  
    --adjust  
    --mperm 10000
```

Empirical p-values in plink.assoc.mperm: rs00008 is experiment-wide significant

CHR	SNP	STAT	EMP1	EMP2
1	rs00001	4.015	0.0489	0.4197
1	rs00002	0.07039	0.7965	1
1	rs00004	0.002133	0.9682	1
1	rs00005	0.3656	0.5391	0.9999
1	rs00006	0.7824	0.3678	0.9965
1	rs00007	0.3887	0.5423	0.9998
1	rs00008	13.14	0.0003	0.0031
1	rs00009	7.681	0.006699	0.06269
1	rs00010	0.5109	0.4799	0.9997
1	rs00011	8.647e-05	0.9961	1
1	rs00012	2.995	0.07449	0.6378
1	rs00014	1.856	0.1748	0.8973
1	rs00015	1.849	0.1724	0.8981

Determine pattern of linkage disequilibrium in the region

```
./plink --file cleaned  
--r2
```

**Calculates pairwise LD (r^2) between all SNPs;
by default, only output only pairs with $r^2 > 0.2$,
to file plink.ld**

CHR_A	BP_A	SNP_A	CHR_B	BP_B	SNP_B	R2
1	15820	rs00008	1	18125	rs00009	0.482748
1	28833	rs00014	1	30974	rs00015	0.948877

Genotypic tests of association at rs00008

```
./plink --file cleaned  
--snp rs00008  
--model
```

Calculates allelic, trend, genotypic, dominant and recessive tests: plink.model

CHR	SNP	A1	A2	TEST	AFF	UNAFF	CHISQ	DF	P
1	rs00008	T	G	GENO	19/235/728	14/172/800	13.89	2	0.0009615
1	rs00008	T	G	TREND	273/1691	200/1772	12.86	1	0.0003354
1	rs00008	T	G	ALLELIC	273/1691	200/1772	13.14	1	0.0002883
1	rs00008	T	G	DOM	254/728	186/800	13.89	1	0.0001934
1	rs00008	T	G	REC	19/963	14/972	0.7913	1	0.3737

Trend test uses Cochran-Armitage test

Genotypic tests of association at rs00008, using logistic regression

```
./plink --file cleaned  
--snp rs00008  
--logistic  
--genotypic
```

Reports results in plink.assoc.logistic

CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P
1	rs00008	15820	T	ADD	1968	1.221	1.123	0.2615
1	rs00008	15820	T	DOMDEV	1968	1.229	1.011	0.312
1	rs00008	15820	T	GENO_2DF	1968	NA	13.8	0.001007

Fits a single model $\text{logit}(P) \sim A + D + e$

Coding of genotypes:

Reports three tests:

$$\begin{aligned}H_0 : A &= 0 \\H_0 : D &= 0 \\H_0 : A = D &= 0\end{aligned}$$

$$\begin{aligned}ADD \ (1df) \\DOMDEV \ (1df) \\GENO \ (2df)\end{aligned}$$

	<i>ADD (A)</i>	<i>DOMDEV (D)</i>
<i>GG</i>	<i>0</i>	<i>0</i>
<i>TG</i>	<i>1</i>	<i>1</i>
<i>TT</i>	<i>2</i>	<i>0</i>

Genotypic tests of association at rs00008, alternate parameterization

```
./plink --file cleaned  
    --snp rs00008  
    --logistic  
    --genotypic  
    --hethom
```

Reports results in plink.assoc.logistic

CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P
1	rs00008	15820	T	HOM	1968	1.491	1.123	0.2615
1	rs00008	15820	T	HET	1968	1.501	3.607	0.0003095
1	rs00008	15820	T	GENO_2DF	1968	NA	13.8	0.001007

Fits a single model $\text{logit}(P) \sim TT + TG + e$

Coding of genotypes:

Reports three tests:

$$H_0 : TT = 0$$

HOM (1df)

HOM HET

GG 0 0

$$H_0 : TG = 0$$

HET (1df)

TG 0 1

$$H_0 : TT = TG = 0$$

GENO (2df)

TT 1 0

Test for sex-specific effects, e.g. a male-only analysis

```
./plink --file cleaned  
        --filter-males  
        --assoc
```

Or formally test SNP-by-sex interaction, using logistic model

```
./plink --file cleaned  
        --snp rs00008  
        --logistic  
        --interaction  
        --sex
```

Reports results in plink.assoc.logistic

CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P
1	rs00008	15820	T	ADD	1968	1.321	2.087	0.03685
1	rs00008	15820	T	SEX	1968	0.9723	-0.2762	0.7824
1	rs00008	15820	T	ADDxSEX	1968	1.174	0.8119	0.4169

Both rs00008 and rs00009 are associated $P<0.01$ and are also in moderately high LD with each other. Are these two associations independent?

```
./plink --file cleaned  
        --logistic  
        --condition rs00008
```

Includes genotype at rs00008 as a covariate; results in plink.assoc.logistic

CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P
1	rs00001	0	G	ADD	1919	0.8403	-1.89	0.05878
1	rs00001	0	G	rs00008	1919	1.436	3.621	0.0002939
1	rs00002	2013	C	ADD	1927	1.013	0.1432	0.8862
1	rs00002	2013	C	rs00008	1927	1.433	3.616	0.0002996
...	...							

Often desirable to extract out only the terms for the SNP (ADD)

```
fgrep -w ADD plink.assoc.logistic
```

1	rs00001	0	G	ADD	1919	0.8403	-1.89	0.05878
1	rs00002	2013	C	ADD	1927	1.013	0.1432	0.8862
1	rs00004	6473	G	ADD	1927	1.015	0.2337	0.8152
1	rs00005	8887	C	ADD	1923	0.9542	-0.7148	0.4747
1	rs00006	11054	T	ADD	1930	1.092	1.012	0.3114
1	rs00007	13413	G	ADD	1922	0.8844	-0.7346	0.4626
1	rs00008	15820	T	ADD	1968	NA	NA	NA
1	rs00009	18125	T	ADD	1936	1.051	0.4677	0.64
1	rs00010	20253	A	ADD	1917	1.054	0.8027	0.4221
1	rs00011	22633	C	ADD	1934	0.9934	-0.08065	0.9357
1	rs00012	24739	C	ADD	1916	1.182	1.793	0.07298
1	rs00014	28833	A	ADD	1929	0.8983	-1.33	0.1834
1	rs00015	30974	A	ADD	1921	0.8954	-1.386	0.1656

```
./plink --file cleaned
--logistic
--condition rs00009
```

Includes genotype at rs00009 as a covariate; results in plink.assoc.logistic

CHR	SNP	BP	A1	TEST	NMISS	OR	STAT	P
1	rs00001	0	G	ADD	1919	0.8541	-1.707	0.08775
1	rs00002	2013	C	ADD	1926	1.068	0.7016	0.4829
1	rs00004	6473	G	ADD	1929	1.01	0.1552	0.8767
1	rs00005	8887	C	ADD	1923	0.9601	-0.6208	0.5347
1	rs00006	11054	T	ADD	1930	1.065	0.7292	0.4659
1	rs00007	13413	G	ADD	1923	0.8862	-0.728	0.4666
1	rs00008	15820	T	ADD	1936	1.349	2.19	0.02854
1	rs00009	18125	T	ADD	1968	NA	NA	NA
1	rs00010	20253	A	ADD	1917	1.041	0.6117	0.5407
1	rs00011	22633	C	ADD	1934	0.9656	-0.4269	0.6695
1	rs00012	24739	C	ADD	1914	1.171	1.693	0.0905
1	rs00014	28833	A	ADD	1929	0.8848	-1.522	0.128
1	rs00015	30974	A	ADD	1921	0.8867	-1.513	0.1304

Given these are in high LD, often useful to explicitly model the haplotypic associations instead

```
./plink --file cleaned  
        --chap  
        --hap-snps rs00008,rs00009
```

```
+++ PLINK conditional haplotype test results +++  
  
2 SNPs, and 3 common haplotypes ( MHF >= 0.01 ) from 4 possible  
  
CHR          BP           SNP     A1      A2          F  
1            15820        rs00008    T       G       0.1202  
1            18125        rs00009    T       C       0.2208
```

Haplogrouping: each {set} allowed a unique effect

Alternate model

{ TT } { GT } { GC }

Null model

{ TT, GT, GC }

HAPLO	FREQ	OR(A)	OR(N)
-----	-----	-----	-----
TT	0.1206	(-ref-)	(-ref-)
GT	0.09969	0.7412	
GC	0.7797	0.705	
-----	-----	-----	-----

Model comparison test statistics:

	Alternate	Null
-2LL :	2671	2684

Likelihood ratio test: chi-square = 12.44
df = 2
p = 0.001992

The --chap command means “conditional haplotype tests”. Output is written to plink.chap

Test rs00008 against haplotypic background

```
./plink --file cleaned  
        --chap  
        --hap-snps rs00008,rs00009  
        --independent-effect rs00008
```

```
Haplogrouping: each {set} allowed a unique effect  
Alternate model  
  { TT }  { GT }  { GC }  
Null model  
  { TT, GT }  { GC }
```

HAPLO	FREQ	OR(A)	OR(N)
TT	0.1206	(-ref-)	(-ref-)
GT	0.09969	0.7412	
GC	0.7797	0.705	0.8086

```
Model comparison test statistics:
```

	Alternate	Null
-2LL :	2671	2676

```
Likelihood ratio test: chi-square = 4.815  
df = 1  
p = 0.02821
```

Test rs00009 against haplotypic background

```
./plink --file cleaned  
        --chap  
        --hap-snps rs00008,rs00009  
        --independent-effect rs00009
```

```
Alternate model  
  { TT }  { GT }  { GC }  
Null model  
  { TT }  { GT, GC }  
  
HAPLO      FREQ      OR(A)      OR(N)  
-----  -----  -----  -----  
  TT      0.1206    (-ref-)    (-ref-)  
  
  GT      0.09969     0.7412     0.7092  
  GC      0.7797      0.705      |  
-----  -----  -----  -----
```

Model comparison test statistics:

	Alternate	Null
-2LL :	2671	2672

Likelihood ratio test: chi-square = 0.2187
df = 1
p = 0.64

Output in Haplovie-friendly format, to confirm LD structure

```
./plink --file cleaned  
--recodeHV
```

Produces two files that can be loaded into Haplovie: plink.ped

```
per0 per0 0 0 1 2 C C A A C C C T A A A A G G T C A A G G C C G G C C  
per1 per1 0 0 1 2 C C C A G C T T A A A A T G T C A C G G T T G G C C  
per2 per2 0 0 2 2 G G C A C C C C A A A A G G C C A C C G T T G G C C  
per3 per3 0 0 1 2 C C A A G G C T T A A A A G G C C A C G G C C G G C C  
per4 per4 0 0 2 2 C C A A C C C C T A A A A G G C C A C C G T T G G C C  
per5 per5 0 0 1 2 C C A A G G T T A A A A T G 0 0 A C C G T T A G A C  
per6 per6 0 0 1 2 C C A A G C C C A A G A T G T C A C C G C T G G C C
```

and plink.info

```
rs00001 0  
rs00002 2013  
rs00004 6473  
rs00005 8887  
rs00006 11054  
rs00007 13413  
rs00008 15820  
rs00009 18125  
rs00010 20253  
rs00011 22633  
rs00012 24739  
rs00014 28833  
rs00015 30974
```

Output in R-friendly format, to confirm SNP-by-sex analysis

```
./plink --file cleaned  
--recodeA
```

Produces a single file that can be loaded into R: plink.raw

FID	IID	PAT	MAT	SEX	PHENOTYPE	rs00008_T
per0	per0	0	0	1	2	0
per1	per1	0	0	1	2	1
per2	per2	0	0	2	2	0
per3	per3	0	0	1	2	0
per4	per4	0	0	2	2	0
per5	per5	0	0	1	2	1
per6	per6	0	0	1	2	1
per7	per7	0	0	2	2	NA
...	...					

```
R  
d <- read.table("plink.raw",header=T)  
str(d)
```

Output in R-friendly format, to confirm SNP-by-sex analysis

```
summary(glm( PHENOTYPE-1 ~ rs00008_T * SEX ,  
           data=d , family="binomial" ) )
```

Same result for interaction test ($P = 0.4169$) as PLINK

Coefficients:					
	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	-0.1308	0.1627	-0.804	0.4212	
rs00008_T	0.5999	0.3220	1.863	0.0624	.
SEX	0.0281	0.1017	0.276	0.7824	
rs00008_T:SEX	-0.1608	0.1981	-0.812	0.4169	

**Note effect arbitrary coding of sex term (M/F):
in PLINK (0/1) versus (1/2) here**

```
summary(glm( PHENOTYPE-1 ~ rs00008_T * I(SEX==1) ,  
           data=d , family="binomial" ) )
```

Coefficients:					
	Estimate	Std. Error	z value	Pr(> z)	
(Intercept)	-0.07465	0.07056	-1.058	0.2901	
rs00008_T	0.27827	0.13331	2.087	0.0369 *	
I(SEX == 1)TRUE	-0.02810	0.10174	-0.276	0.7824	
rs00008_T:I(SEX == 1)TRUE	0.16084	0.19811	0.812	0.4169	

In summary

- We have performed basic QC and association analysis on a candidate gene case/control dataset
- The SNP rs00008 showed a significant association ($P=3\times 10^{-4}$) and was significant after correction for multiple testing by permutation ($P=0.003$)
- The T (versus G) allele has a 12% sample frequency and an allelic odds ratio of 1.43
- An additive model fits the data well versus a 2df genotypic model ($P=0.31$ for genotypic vs allelic)
- There is no indication of sex-specific effects ($P=0.42$)
- Haplotype-based tests shows the weaker association at nearby rs00009 does not represent an independent signal

Questions?